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In the Specification:

Please replace the sequence listing filed on October 11, 2006 with the attached sequence listing.

Please amend the specification on page 23 lines 9-13 as follows:

Sequence of *Lin ext ba*:

5'-GAT GCC GGC CAC GAT GCG TCC GGC-3' (SEQ ID NO. 1)

Sequence of *Hae sub fo*:

5'-C GTC ATG **GFC** TAT GCG GGC GAC CAC ACC CGT CCT GTG GAT-3'
(SEQ ID NO. 2)

Please amend the specification on page 27 carrying over onto page 28 as follows:

For the expression of the proteins and the production of the covalent protein-DNA complexes the samples were incubated at 30 °C for 150 min. Subsequently, the aqueous phase containing the DNA-protein fusions was extracted from the emulsion as follows:

The samples were centrifuged for 10 min. at ~~7,000~~ 7,000 rpm, whereafter the water compartments sedimented at the bottom of the reaction vials. The supernatant (oil phase) was suctioned off and 150 µl buffer were added (buffer consisting of: TBS (Tris-buffered saline) with 1 mM CaCl₂ (=TBSC), pH 7.4, 5 µM biotinylated double-stranded DNA fragments for blocking the magnetic beads employed later on [5'-biotin-GGA GCT TCT GCA TTC TGT GTG

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CTG-3' (SEQ ID NO. 3) (Qiagen)], 1 μ M competing double-stranded DNA fragments [5'-ATC TAA **GGC** CAA TGT ACT AGA **CGG** CCA TTC CAG ATG CAG **GCC** AAG CGT ACA TAC **GGC** CTA GCT AAA TCA AGG **CCG** TAT CGT-3' (SEQ ID NO. 4), substrate sequence for *M.Hae III* in bold letters (Qiagen)] followed by 1 ml diethyl ether. Subsequently, the sample was shaken with a vortex for 2 x 10 sec. After the separation of the water phase and the oil phase the aqueous phase lying below was removed with a pipette and dried in a 24-microtiter plate for 10 min., so that the remaining diethyl ether was allowed to evaporate completely.

During the extraction of the aqueous phase 25 μ l magnetic beads coated with streptavidine (Dynabeads, Dynal, Norway) were incubated with biotinylated, Calmodulin-binding peptide (400 nM, biotin-CAAARWKKAFIAVSAANRFKKIS (SEQ ID NO. 5) (Montigiani et al., 1996) or with biotinylated anti-Flag antibody M2 (2 μ l/50 μ l beads, M2 antibody, Sigma-Aldrich) for 15 min. The Calmodulin-binding peptide was used to select the *M.Hae III*-Calmodulin-DNA fusions located in the aqueous phase of the emulsion, whereas the anti-Flag antibody was employed as a negative control. After the incubation of the magnetic beads with peptides or antibodies these were washed once with TBSC 0.1% Tween 20 (Fluka). Subsequently the beads were blocked for 15 min. at room temperature with biotinylated DNA fragments (5 μ M) [5'-biotin-GGA GCT TCT GCA TTC TGT GTG CTG-3' (SEQ ID NO. 3) (Qiagen)].

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carrying over onto page 30 as follows:

To the transcription/translation mixture a mixture of 10^9 DNA molecules in total was added, wherein a factor of 4200 more DNA molecules coded for the fusion protein *M.Hae III*-ED-B than for *M.Hae III*-Calmodulin. The selection experiment was done with magnetic beads that had been coated either with Calmodulin-binding peptides or with anti-Flag antibodies (M2, Sigma-Aldrich). The result of the experiment was evaluated by real-time PCR.

However, the magnetic beads were not used for the real-time PCR directly, but the selected DNA molecules were first amplified in a PCT with the primers Ampl ba (5'-CCC GCG AAA TTA ATA CGA CTC A-3', (SEQ ID NO. 6) Qiagen) and Ampl fo (5'-AAA ACC CCT CAA GAC CCG TT-3', (SEQ ID NO. 7) Qiagen). The PRC was performed with the following temperature program:

94 °C (3 min.) → [94 °C (45 sec.) → 51 °C (1 min.) → 72 °C (100 sec.)]_{35 cycles} → 72 °C (3 min.) → 4 °C.